

CelCradle-500 Technical Report XIII Cultivation of CHO-K1 Cells and Production of β-Gal

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1 Description

CelCradle-500 provides a powerful tool to achieve high cell density and high productivity of target bioproducts in a cell culture because it has a unique feature of offering high oxygen transfer and low shear stress culture environment. Users can easily collect highly concentrated cells, virus or secreted products from one 500 ml CelCradle-500 bottle. In this study, the application of CelCradle-500 beta-gal is illustrated. 5 x 10⁷ cells/bottle was seeded and obtained a total of 1.4 x 10⁹ cells counted by crystal violet dye nucleus count method at 234.5 hours, with a total 28 folds increase of cell population. Beta-gal total yield is equivalent to the production from $320 \times T-150$ flasks or 56.5 x Roller bottles. Beta-gal productivity per cell is 3.82 folds increased compared with the cells in T-150 flask. This technical sheet provides a general protocol for users to start up their culture. However, the optimum condition of each cell culture for each case may require the users to determine.

2 Material

Device	Cell Line	Medium	Seed
CelCradle-500	CHO-β-gal	DMEM/F12/5%FBS + 2.5 g/L glucose + 2.5 mM glutamine + 3.7 g NaHCO ₃	5 x 10 ⁷ cells/bottle

3 Protocol

3.1 Inoculum Preparation

Prepare 3 x T-150 Flasks. Seed with 3 x 10^6 cells each. Culture at 37°C, 5% CO2 for a total of 3 days. Replenish medium at day 3. Harvest cells by standard trypsinization protocol. Prepare 5 × 10^7 suspended cells with viability above 95%, and concentrate cells in 50 ml culture medium.

3.2 Inoculation

Pre-warm DMEM/10%FBS medium in 37°C water bath. Take out one CelCradle-500 bottle aseptically and place in a biosafety cabinet. Open the cap and add 450 ml culture medium in the bottle. Well distribute 30 ml prepared inoculums on top of the matrix box and bring to CelCradle Stage immediately. Fix the bottles on CelCradle Stage controller in CO₂ incubator with 37°C, and 5% CO₂ and start the run immediately.



3.3 Immobilization

Set up operation parameters on the CelCradle Stage control box and start the controller by pressing "START" button. The inoculation parameters are set as below:

Rising rate	Top Holding Time	Down Rate	Bottom Holding Time
2.0 mm/s	20 sec	2.0 mm/s	0 sec

3.4 Culture

After 3.5 hours, switch the parameters to culture parameters. The culture control parameters are set as below:

Rising rate	Top Holding Time	Down Rate	Bottom Holding Time
1.5 mm/s	0 sec	1.5 mm/s	1 min 30 sec

Monitor the residual glucose concentration and the color of medium in order to predict the time to change culture medium.

The setup parameters are only for reference. These are not necessarily the optimum parameter conditions.

4 Result

Glucose and Lactate profile





Glutamine and Ammonia profile



Glucose uptake rate (GUR) and Lactate production rate (LPR) profile



Glutamine uptake rate (GnUR) and Ammonia production rate (APR) profile





L/G and A/Gn



Cell grow curve by crystal violet dye nuclear count method



Cell propagation is exponentially increased until 144 hours. There is an inhibition of cell growth after 144 hours and may due to the low pH within that period.

pH/CO₂





pH is below normal range after 144 hours culture. Cell growth is inhibited thereafter. We will suggest to reduce initial glucose concentration to below 3.0 g/L and increase NaHCO₃ to above 4.5 g/L and also adjust pH twice a day if the low pH situation occurred.

β-gal production



Comparison between CelCradle-500, and T-150

	CelCradle	T-150
Culture Period (day)	10	3
Cell Mass (cells)	1.40 x 10 ⁹	1.67 x 10 ⁷
Productivity (OD420/10 ⁷ cells)	2.6	0.68
Total Yield (OD420)	364	1.136
Equivalent to T-150	320	1
Equivalent to Roller Bottle	56.5	0.18

The growth of CHO-k1 cells in CelCradle-500 is very fast and require lower seeding density, it took 6 days to reach nearly 28 folds increase of cell population, and reach 1.4×10^9 cells/bottle. However, the cell ceased to grow after 6 days probably due to the sever fluctuation of pH during culture. GUR is very high and produce a lot of lactate. PH is below normal range during late phase of culture and results an inhibition of cell growth. We suggest to reduce the initial glucose concentration to below 3 g/L and gradually increase NaHCO₃ to above 4.5 g/L in order to control the pH within range.



5 Summary

Seed	Inoculum Volume	Medium Volume	Medium
5 x 10 ⁵ cells/bottle	50 ml	500 ml	DMEM/5%FCS
Total Culture Age	Total Medium Consumed	Total Medium Replenish Frequency	Final Cell Density (Nuclei Count)
244 hours	3720 ml	7 times	1.40 x 10 ⁹ cells/bottle

6 VacciXcell Technical Support

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